

Il ruolo di EFL nella gestione della qualità nel laboratorio di Istocompatibilità e Immunogenetica

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EFI Mission

Mission and goals

It is the mission and goal of EFI:

- 1 To support the development of Immunogenetics in Europe as a discipline in medicine and promote research and training in this field.
- 2 To provide a forum for exchange of scientific information and to reinforce the skills and knowledge of young scientists and others working in the field.
- 3 To create a formal organisation of workers in the field of immunogenetics, histocompatibility testing and transplantation.
- 4 To develop recommendations for standardisation of techniques, quality control and criteria for accreditation and to support their implementation.
- 5 To promote the organisation and use of immunogenetic databases.
- 6 To develop relations with organisations with similar aims.

The association shall abstain from any type of political activity.

Sviluppare in Europa l'Immunogenetica, promuovendo la ricerca e la formazione in questo campo

Promuovere scambi di informazioni scientifiche e promuovere la conoscenza sia di giovani che di altri nel settore dell'immunogenetica

Sviluppare raccomandazioni per la standardizzazione delle tecniche di laboratorio, della gestione della qualità, dei criteri utili al supporto del processo di accreditamento

Consolidare le relazioni con organizzazioni mondiali simili ad EFI

Committees

Executive Committee

Accreditation Committee

Education Committee

EPT Committee

IT & Bioinformatics Committee

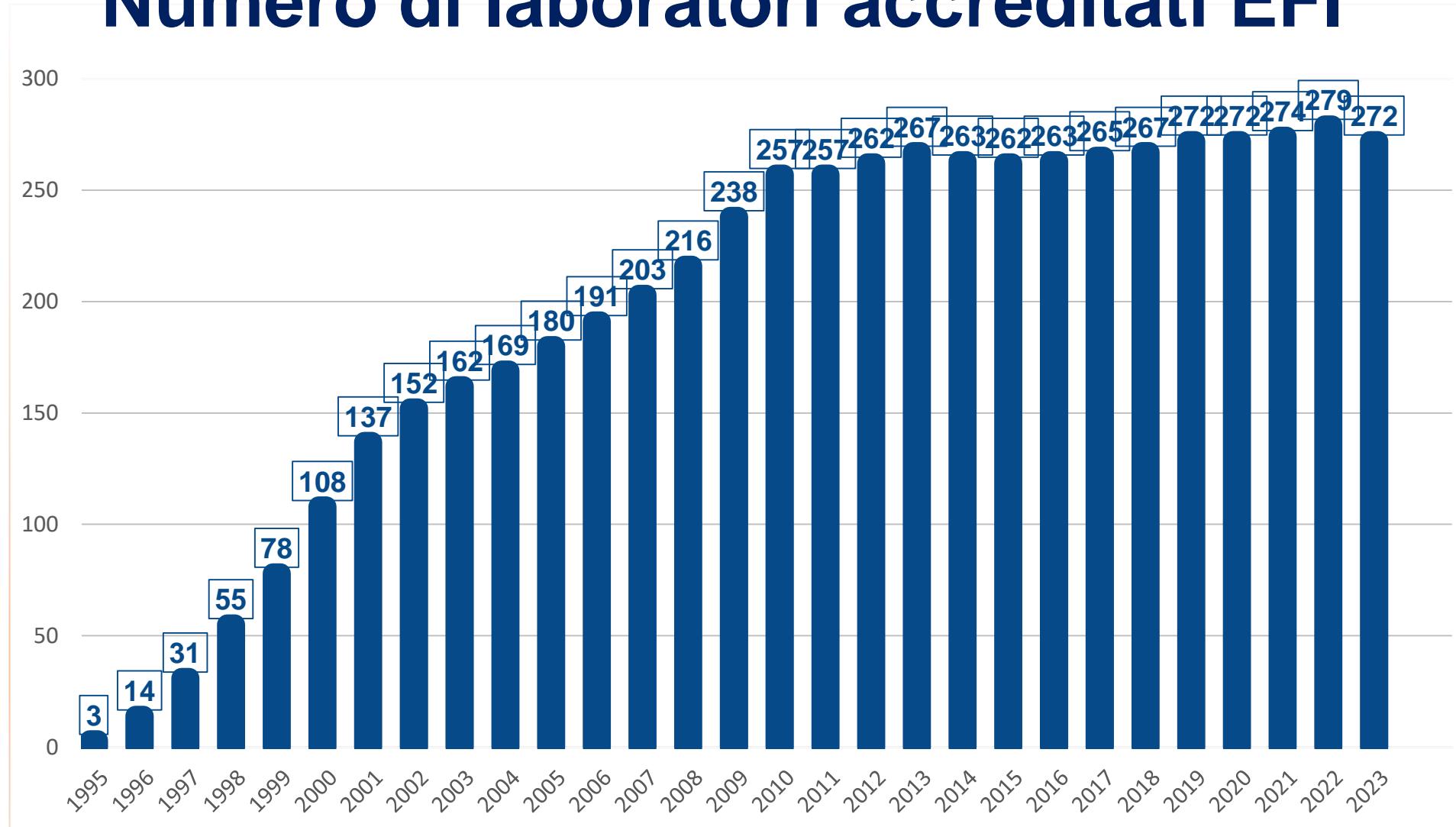
Scientific Committee

Accreditation Committee

Promuovere il Sistema Qualità nell'attività di trapianti di
organi solidi e cellule staminali ematopoietiche

Rilascio di una Certificazione di Accreditamento

Numero di laboratori accreditati EFI



Distribuzione dei laboratori accreditati EFI nel mondo nel 2023

REGION	ACCREDITED LABORATORIES	
01	SCANDINAVIA	12
02	BENELUX	12
03	UK + IRELAND	20
04	GERMANY	40
05	CENTRAL EUROPE	20
06+11	FRANCE + SWITZERLAND	33
07	ITALY	50
08	SE EUROPE, ISRAËL, ARMENIA	28
09+10	SPAIN + PORTUGAL	26
99	SOUTH AFRICA, CHINA, KUWAIT, COLOMBIA, USA, INDIA, ARGENTINA, BRAZIL,	12
99	COMMISSIONERS LABS	19
	TOTAL	272

Istituti internazionali che richiedono l'accreditamento EFi o ASHI

WMDA (World Marrow Donor Association)

NMDP (National Marrow Donor Program)

JACIE (Joint Accreditation Committee of ISCT-Europe and EBMT)

FACT (Foundation for the Accreditation of Cellular Therapy)

NETCORD FAHCT (Cord Blood Sharing Organization)

BSBMT/UKCCSG (British Society for Bone Marrow Transplantation)

IBMDR (Italian Bone Marrow Donor Registry)

EUROTRANSPLANT (Organ Sharing Organization)

Napoli, 10/12 ottobre 2024

Il percorso di accreditamento EFI

Categorie cliniche e tecniche

- **Clinical categories**

- Renal Transplantation and Other Solid Organ Transplant
- Haematopoietic Transplant
- Disease Association
- Blood Transfusion

- **Technical categories**

- HLA Typing
- Crossmatching
- Antibody screening & identification

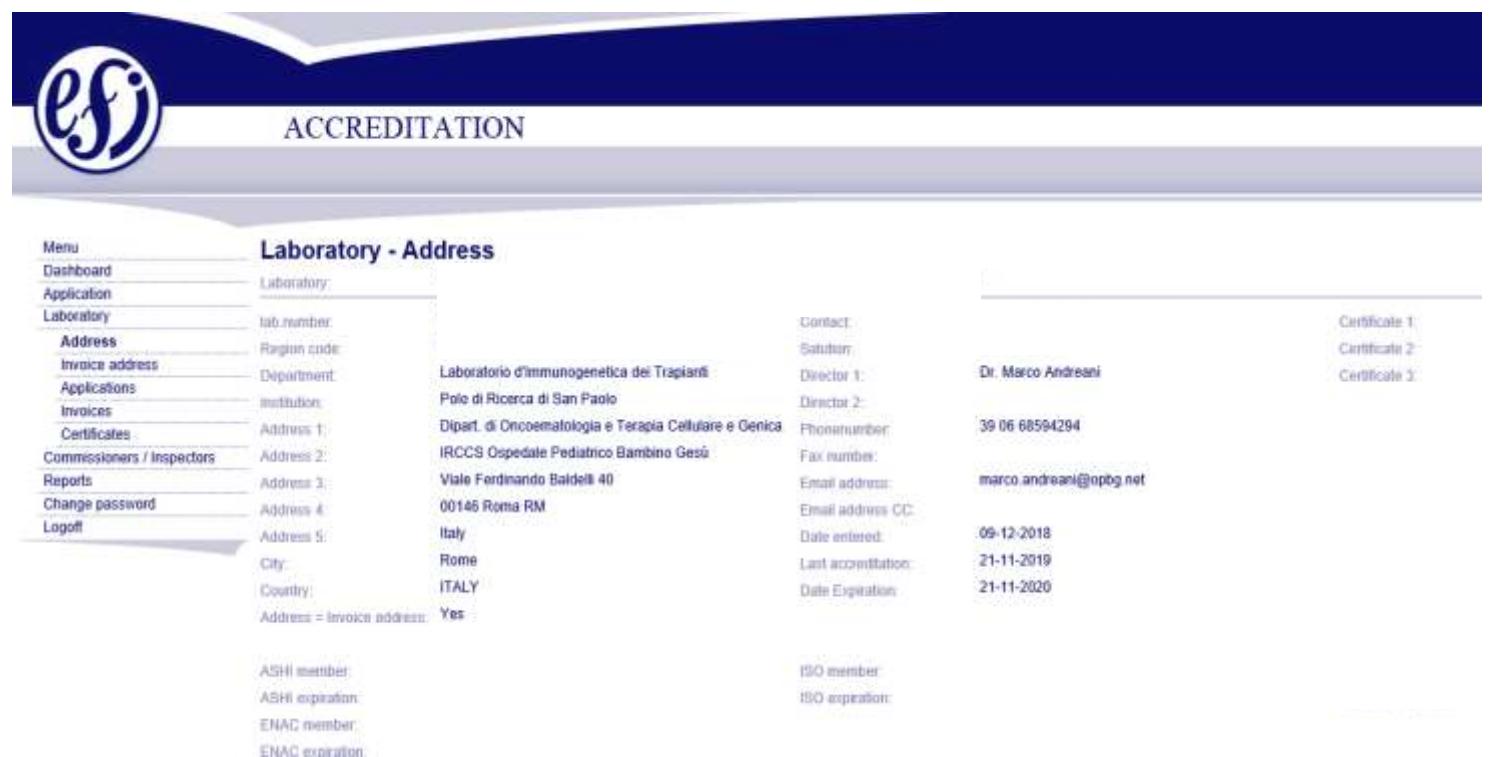
Categories	Minimum Requirements	Notes
Renal and/or Pancreatic Transplantation		
Recipient Typing	HLA Class I and II low resolution (1st field)	A,B,DR + other loci required by national regulations
Antibody Screening	Antibody detection HLA Class I and Class II	
Antibody Identification	Antibody specificity HLA Class I and Class II	
Donor Typing	HLA Class I and II low resolution (1st field)	
Crossmatching	Antibody specificity and Crossmatching.	If Virtual Crossmatching is used for any sensitised patients specificity with single antigen beads required
Other Solid Organ Transplantation		
Recipient Typing	For sensitized patients	
Antibody Screening	Antibody detection HLA Class I and Class II	
Antibody Identification	Mandatory if screening is positive	
Donor Typing	For sensitized recipients	
Crossmatching	For sensitized recipients	

Categories	Minimum Requirements	Notes
Haematopoietic Stem Cell Transplantation		
Donor Registry Typing	HLA Class I and Class II typing low resolution (1 st field)	
Related Transplantation	HLA Class I and Class II typing high resolution (2 nd field)	As required to determine identity
Unrelated Transplantation	HLA Class I and Class II typing high resolution (2 nd field)	A,B,C,DRB1 + additional loci if required by transplant protocol
Cord Blood Typing	HLA Class I and Class II low resolution	
Crossmatching	Crossmatching or antibody specificity	For haplotype-identical Tx (currently recommended, to become mandatory in future version of standards)
Chimaerism and Engraftment Monitoring	As required by the transplant protocol	
Disease Association Studies	HLA Class I and Class II typing low resolution and high resolution as required	Where an association is with specific alleles typing to 2 nd field is required
Transfusion	HLA/HPA/HNA antibody screening and typing as required	

In cosa consiste il processo di Accreditamento EFI

On-line submission

- Application form
- Multiple supporting documents



The screenshot shows a web-based application for laboratory accreditation. The top navigation bar includes the 'efi' logo and the word 'ACCREDITATION'. The left sidebar lists various menu items: Menu, Dashboard, Application, Laboratory, Address, Invoice address, Applications, Invoices, Certificates, Commissioners / Inspectors, Reports, Change password, and Logoff. The main content area is titled 'Laboratory - Address'. It contains two columns of input fields. The left column includes: Lab number, Region code, Department, Institution, Address 1, Address 2, Address 3, Address 4, Address 5, City, Country, and Address = Invoice address (with a 'Yes' checkbox). The right column includes: Contact, Surname, Director 1, Director 2, Phone number, Fax number, Email address, Email address CC, Date entered, Last accreditation, Date expiration, Certificate 1, Certificate 2, and Certificate 3. At the bottom, there are additional fields for ASHI member, ASHI expiration, ENAC member, and ENAC expiration.

Left Column Fields	Right Column Fields
Lab number	Contact
Region code	Surname
Department	Director 1
Institution	Director 2
Address 1	Phone number
Address 2	Fax number
Address 3	Email address
Address 4	Email address CC
Address 5	Date entered
City	Last accreditation
Country	Date expiration
Address = Invoice address: Yes	Certificate 1
	Certificate 2
	Certificate 3
ASHI member	ISO member
ASHI expiration	ISO expiration
ENAC member	
ENAC expiration	

In cosa consiste il processo di Accreditamento EFI

Application

EFI Accreditation Program

EFI No.

Application file accreditation EFI

Please fill in: Application A Application C Date:

Application is filled in according to the Standards version 8.0

Laboratory/institute name:

To complete the application A/C, we kindly ask you to fill in all the sections of the application file (indicate any section which are not applicable). Also please complete the separate sections for the address data and names as they should appear on the certificate and for the accreditation 'Categories', you will find both sections on the Accreditation website.

Submit an organogram of the laboratory with positions and names of persons at the staff and supervisory levels including the quality manager (addendum #1). If the laboratory is part of a larger department, an overview of the department must also be provided (addendum #2). Indicate the position of the quality manager on addendum #1 and/or #2 as appropriate.

We do hereby apply to EFI for laboratory accreditation in the area(s) indicated at the Accreditation website

It is understood that granting of accreditation is dependent on compliance with all applicable EFI Standards. If a conflict of interest exists with any individual involved in the accreditation procedure we will bring that to the notice of the Chairperson of the EFI Accreditation Committee before the inspection takes place.

The Directors confirm that all information is truthful and accurate to the best of our knowledge.

I/we consent to the storage and use of the data submitted in this application for the purposes of EFI accreditation.

PERSONNEL

Director/Co-Director Qualifications				
Last name	First name	What is the average hours spent in the laboratory per week.	Is emergency consultation available during your absence?	Weeks/year away from institution? *
			Yes	
			N/A	

*) for periods of > 3 consecutive workdays

Describe in an addendum (addendum #3) your duties in your present position, especially your role in the laboratory, including the extent to which you participate in the review, interpretation and reporting of test results, development and performance or supervision of test procedures, training and evaluation of staff and fellows and establishment of laboratory policies. Include a policy defining who may act as a designated individual for signing reports. Confirm that you are aware of the relevant national legislation.

Submit a complete Curriculum Vitae (addendum #4). This CV must also include the degrees earned, training received, length of time in present position.

Submit a list of publications (addendum #5).

Note: in case of change in directorship of a laboratory the commissioner must be informed immediately and no later than 7 days following the change, a CV of the new director must be provided.

Additional Documents

EFI Accreditation Program

EFI No.

List of documents to be submitted separately as addenda: Please note: all addenda except reports and SOPs must be written in English.

Addendum	Document	Submitted
# 1	Laboratory's organogram with names /positions of staff/supervisors	Yes <input type="checkbox"/>
# 2	Organogram of the department/institute indicating the laboratory's position within, and the position of the Quality Manager if applicable	Yes <input type="checkbox"/>
# 3	Overview of duties of the director(s), co-director(s) and technical supervisor(s)	Yes <input type="checkbox"/>
# 4	Complete C.V.'s of director(s), co-director(s) and technical supervisor(s)	Yes <input type="checkbox"/>
# 5	List of publications of director(s), co-director(s) and technical supervisor(s)	Yes <input type="checkbox"/>
# 6	Continuing education of all histocompatibility technical staff (section III A)	Yes <input type="checkbox"/>
# 7	Modifications in the Laboratory (Not applicable to application A)	Yes <input type="checkbox"/>
# 8	Details of antibody screening/identification testing	Yes <input type="checkbox"/>
# 9	Details of crossmatch testing	Yes <input type="checkbox"/>
# 10	Overview molecular biology techniques for each locus	Yes <input type="checkbox"/>
# 11	Protocols for renal and/or pancreatic transplantation	No <input type="checkbox"/>
# 12	Protocols for other solid organ transplantation	No <input type="checkbox"/>
# 13	Local clinical protocol related HSCT	Yes <input type="checkbox"/>
# 14	Copy of certificate subcontracted laboratory	No <input type="checkbox"/>
# 15	Anonymous copy of selected case for related HSCT	Yes <input type="checkbox"/>
# 16	Local clinical protocol unrelated HSCT	Yes <input type="checkbox"/>
# 17	Anonymous copy of selected case for unrelated HSCT	Yes <input type="checkbox"/>
# 18	Overview of techniques used chimerism	Yes <input type="checkbox"/>
# 19	Overview of techniques used for transfusion testing	No <input type="checkbox"/>
# 20	List of worksheets and prints of screens for data storage (if applicable)	Yes <input type="checkbox"/>
# 21	List of content procedure manual/SOPs	Yes <input type="checkbox"/>
# 22	Proficiency test results, including consensus report and certificate	Yes <input type="checkbox"/>
# 23	SOP for competence evaluation	Yes <input type="checkbox"/>
# 24	List of content of the QA Manual, if applicable	Yes <input type="checkbox"/>
# 25	Result of internal audit by QA Officer, if applicable	Yes <input type="checkbox"/>
# 26	Documentation that the laboratory is in accordance with relevant national legislation	Yes <input type="checkbox"/>

After inspection:

EFI-IQ: Completed inspection questionnaire at: http://www.surveymonkey.com/efiinspection_questionnaire

In cosa consiste il processo di Accreditamento EFI

Tutti i documenti sono inizialmente valutati dal Commissario EFI (3 in Italia)

- Dottor Franco Papola
- Dottoressa Annamaria Pasi
- Dottoressa Elena Longhi

Successivamente il Commissario sceglierà gli Ispettori, che valuteranno la documentazione, prima della site visit

Region	Position	Last name	First name	City	Country
7a	Commissioner	Papola	Franco	L'Aquila	Italy
7b	Commissioner	Longhi	Elena	Milano	Italy
7c	Commissioner	Pasi	Annamaria	Pavia	Italy
7	Inspector	Battarra	Mariarosa	Rome	Italy
7	Inspector	Curcio	Michele	Pisa	Italy
7	Inspector	Ferrarese	Diego	Piacenza	Italy
7	Inspector	Garbarino	Lucia	Genova	Italy
7	Inspector	Laurenti	Luca	Rome	Italy
7	Inspector	Troiano	Maria	Rome	Italy
7	Inspector	Vecchiato	Cinzia	Bolzano	Italy
7	Inspector	Mazzi	Benedetta	Milano	Italy
7	Inspector*	Andreani	Marco	Rome	Italy
7	New Inspector	Scarpa	Alice	Piacenza	Italy

In cosa consiste il processo di Accreditamento EFI

3 year cycle of accreditation

Year 0: on-site inspection after lab Application(Packet A)

Year 1: & 2 document review + self inspection (Packet B1 & B2)

Year 3: on-site inspection (Packet C)

In cosa consiste il processo di Accreditamento EFI

- A - site visit di un giorno eseguita da 2 ispettori (1 locale, 1 straniero)**
 - Valutazione dei locali, delle attività, degli strumenti, oltre che delle procedure e di qualsiasi documento inerente la richiesta di accreditamento
 - Compilazione della check list
 - Invio del report al Commissario
- B - Il Commissario, dopo aver esaminato il report degli Ispettori**
 - Invia al Laboratorio l'elenco delle non conformità rilevate e la richiesta (se necessario) di azioni correttive
- C - Il Laboratorio invia al Commissario una comunicazione con le informazioni legate alla risoluzione delle non conformità**
- D - Il Commissario dispone il rilascio della certificazione di Accreditamento**

Napoli, 10/12 ottobre 2024

DOCUMENTI

EFI Standards

European Federation for Immunogenetics



**STANDARDS FOR
HISTOCOMPATIBILITY
& IMMUNOGENETICS
TESTING**

Version 8.0

Approved by the Standards and Quality Assurance Committee on 10th May 2016
Accredited by the EFI Executive Committee on 29th August 2016
(Effective from January 1st 2017)
(Version 8.0)

SECTION A – GENERAL POLICIES

SECTION B – PERSONNEL QUALIFICATIONS

SECTION C – QUALITY ASSURANCE

SECTION D – EXTERNAL PROFICIENCY TESTING

SECTION E – ANALYSIS PROCESSES

SECTION F – POST-ANALYSIS PROCESSES

EFI STANDARDS

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EFI Standard	efi	Not Applic. Yes	Required Yes No	Recom.* Yes No
SECTION B – PERSONNEL QUALIFICATIONS				
B1	For the purposes of this document, EFI defines the Director as the person who is responsible for the H&I laboratory activities for which accreditation is applied for			
B2	The laboratory must employ one or more individuals who meet the qualifications and fulfil the responsibilities of:			
B3	The Director and/or Co-Director			
B3.1	A Director, that must:			
B3.1.1	Hold a qualification approved by EFI, such as an ESHI or national diploma, earned doctoral degree in a biological science, or be a physician, and	<input type="checkbox"/>	<input type="checkbox"/>	
B3.1.2	Have minimum qualification experience equivalent to either of the following:			
B3.1.2.1	Four years' relevant experience two of which were devoted to full time training in human H&I testing, or	<input type="checkbox"/>	<input type="checkbox"/>	
B3.1.2.2	Four years of working experience at full time in human H&I testing	<input type="checkbox"/>	<input type="checkbox"/>	
B3.1.2.3	Additional qualifications required according to national legislation also apply	<input type="checkbox"/>	<input type="checkbox"/>	
B3.1.2.4	If these tests are performed in a laboratory not performing H&I. For chimaerism, KIR, HPA, HNA two years of working experience at full time	<input type="checkbox"/>	<input type="checkbox"/>	
B3.1.3	Have documentation of professional competence in the appropriate activities in which the laboratory is engaged. This should be based on sound knowledge of the fundamentals of immunology, genetics and histocompatibility testing as appropriate to the areas in which accreditation is sought	<input type="checkbox"/>	<input type="checkbox"/>	
B3.1.4	If a Co-Director is appointed, this person must also fulfil Standards B3.1.1 - B3.1.3	<input type="checkbox"/>	<input type="checkbox"/>	
B3.1.5	The Director or Co-Director must:			
B3.1.5.1	Be available at on site to supervise the laboratory for at least 80% of the week	<input type="checkbox"/>	<input type="checkbox"/>	
B3.1.5.2	Provide adequate supervision of technical personnel	<input type="checkbox"/>	<input type="checkbox"/>	
B3.1.5.3	Utilises his/her special scientific skills in developing new procedures	<input type="checkbox"/>	<input type="checkbox"/>	
B3.1.5.4	Be held responsible for the proper performance, interpretation and reporting of all laboratory procedures	<input type="checkbox"/>	<input type="checkbox"/>	
B3.1.5.5	Ensure the laboratory's successful participation in proficiency testing.	<input type="checkbox"/>	<input type="checkbox"/>	
B3.1.5.6	Be informed of the relevant national legislation and professional standards	<input type="checkbox"/>	<input type="checkbox"/>	
B3.1.5.7	Comply with the relevant national legislation and professional standards	<input type="checkbox"/>	<input type="checkbox"/>	
B3.1.5.8	Demonstrate active participation in clinically relevant professional development, such as national or international conferences or workshops	<input type="checkbox"/>	<input type="checkbox"/>	
B3.1.6	The Director or Co-Director should:			
B3.1.6.1	Have publications in peer-reviewed journals			<input type="checkbox"/>
B4	Technical Staff			
B4.1	A Technical Supervisor, that must:			

Alcuni esempi di gestione della qualità

- **Caratteristiche delle aree di lavoro**
- **Processo di accettazione dei campioni biologici**
- **Appropriatezza della documentazione**
- **Appropriata manutenzione delle attrezzature**
- **Controlli di qualità esterni**
- **Qualifica del Personale**
- **Validazione**

Alcuni esempi di gestione della qualità

- **Caratteristiche delle aree di lavoro**
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 - Validazione

Caratteristiche delle aree di lavoro

C2.1.3	Laboratories performing amplification of nucleic acids must use:
C2.1.3.1	A dedicated work area with restricted traffic flow
C2.1.3.2	Physical and/or biochemical barriers to prevent DNA contamination, including the use of dedicated
C2.1.3.2.1	Equipment
C2.1.3.2.2	Laboratory coats
C2.1.3.2.3	Disposable supplies
C2.1.4	Pre-amplification procedures must be performed in an area which excludes amplified DNA that has the potential to serve as a template for amplification in any of the genetic systems tested in the laboratory
C2.1.5	All activities occurring from and including thermal cycling must take place in the post-amplification area

I laboratori che eseguono amplificazione degli acidi nucleici, per prevenire la possibile contaminazione devono disporre di aree specifiche, strumenti, camici etc. dedicati all'area definita di Pre-PCR

Caratteristiche delle aree di lavoro



Caratteristiche delle aree di lavoro

- I frigoriferi e i congelatori che contengono reagenti dedicati ad attività Pre- o Post-PCR devono essere necessariamente posizionati nelle aree specifiche
- I camici del personale che transitano da aree Pre- a Post-PCR devono essere abitualmente cambiati nell'area considerate filtro



Filter Area



Pre-PCR



Post-PCR 1



Post-PCR 2

Caratteristiche delle aree di lavoro

C2	TECHNICAL			
C2.1	Facilities			
C2.1.1	<i>The following facilities must be adequate and immediately available to the laboratory:</i>			
C2.1.1.1	Sufficient space so that all procedures can be carried out without crowding to the extent that errors may result, in accordance with national regulations	<input checked="" type="checkbox"/>	<input type="checkbox"/>	
C2.1.1.2	Lighting	<input checked="" type="checkbox"/>	<input type="checkbox"/>	
C2.1.1.3	Ventilation	<input checked="" type="checkbox"/>	<input type="checkbox"/>	



Alcuni esempi di gestione della qualità

- Caratteristiche delle aree di lavoro
- **Processo di accettazione dei campioni biologici**
- Appropriatezza della documentazione
- Appropriata manutenzione delle attrezzature
- Controlli di qualità esterni
- Qualifica del Personale
- Validazione

Processo di accettazione dei campioni biologici

C3.2	Sample acceptance			
C3.2.1	<i>The laboratory must:</i>			
C3.2.1.1	Maintain a system to ensure reliable specimen identification	<input checked="" type="checkbox"/>	<input type="checkbox"/>	
C3.2.1.2	Document each step in the processing and testing of patient specimens to assure that accurate test results are recorded	<input checked="" type="checkbox"/>	<input type="checkbox"/>	
C3.2.1.3	Have criteria for specimen rejection	<input type="checkbox"/>	<input checked="" type="checkbox"/>	
C3.2.1.4	Have mechanism to assure that specimens are not tested when they do not meet the laboratory's criteria for acceptability	<input type="checkbox"/>	<input checked="" type="checkbox"/>	

No SOP describing the appropriate criteria for specimen rejection is present in the lab



I laboratori devono disporre di un sistema che permetta la corretta identificazione dei campioni biologici, di documentazione che assicuri l'accuratezza dei risultati raccolti, di criteri scritti per eventualmente rigettare un campione biologico etc.

Alcuni esempi di gestione della qualità

- Caratteristiche delle aree di lavoro
- Processo di accettazione dei campioni biologici
- **Appropriatezza della documentazione**
- Appropriate manutenzione delle attrezzature
- Controlli di qualità esterni
- Qualifica del Personale
- Validazione

Liste delle Procedure

Titolo del Documento	ED	REVISIONI				
		0 (data)	1 (data)	2 (data)	3 (data)	4 (data)
PO 22 LIT GESTIONE DELLA QUALITA'	3	30/09/2019	01/10/2020			
PO 23 LIT CONTROLLO DELLE TEMPERATURE	3	30/09/2019	10/03/2020	09/09/2020		
PO 24 LIT MONITORAGGIO DELLA CONTAMINAZIONE	3	30/09/2019	01/10/2020			
PO 25 LIT GESTIONE DI REAGENTI E MATERIALE MONOUSO	3	30/09/2019	05/03/2020	09/09/2020	14/06/2022	
PO 26 LIT PROGRAMMAZIONE DELL'ATTIVITA' E GESTIONE DEI DATI	3	30/09/2019	01/10/2020			
PO 27 LIT TIPIZZAZIONE HLA	3	30/09/2019	01/10/2020			
PO 28 LIT ESTRADONE DEL DNA	3	30/09/2019	30/04/2020	09/09/2020		
PO 29 LIT TIPIZZAZIONE MEDIANTE PCR - SSP	3	30/09/2019	01/10/2020			
PO 30 LIT TIPIZZAZIONE MEDIANTE PCR-SSO LUMINEX	3	30/09/2019	01/10/2020			
PO 31 LIT MONITORAGGIO ATTECCHEMENTO	3	30/09/2019	01/10/2020			
PO 32 LIT TIPIZZAZIONE MEDIANTE PCR - SBT	3	30/09/2019	01/10/2020	dimessa		
PO 33 LIT TIPIZZAZIONE HLA MEDIANTE NGS	3	30/09/2019	01/10/2020	25/09/2020		
PO 34 LIT DETERMINAZIONE E IDENTIFICAZIONE DI ANTICORPI ANTI - HLA	3	30/09/2019	01/10/2020	01/02/2021		
PO 35 LIT ATTIVITA' IMMUNOGENETICA NEL TRAPIANTO DI ORGANO SOLIDO	3	01/10/2020	01/02/2021			
PO 36 LIT CROSMATCH NEL TRAPIANTO DI ORGANO SOLIDO	3	01/10/2020	01/02/2021			
PO 37 LIT STUDIO DEI GENI KIR NELL'ANALISI DEL DONATORE DI CSE	3	01/10/2020				
PO 38 LIT DETERMINAZIONE CITOFUORIMETRICA NELL'ANALISI DI CROSMATCH FC-XME FLOW PRA	3	01/10/2020	01/02/2021			

Liste delle Istruzioni

Titolo del Documento	ED	REVISIONI				
		0 (data)	1 (data)	2 (data)	3 (data)	4 (data)
ISTR 01 PO 24 LIT ESECUZIONE DEI CONTROLLI DI CONTAMINAZIONE	3	30/09/2019				
ISTR 01 PO 25 LIT UTILIZZO DEL SISTEMA MISSION	3	30/09/2019				
ISTR 01 PO 26 LITISTRUZIONE OPERATIVA SOFTWARE GESTIONALE DNLab	3	30/09/2019	09/04/2020			
ISTR 01 PO 27 LIT TIPIZZAZIONE HLA PER LA RICERCA DI UN DONATORE CORRELATO	3	30/09/2019	09/04/2020			
ISTR 01 PO 28 LIT ESTRATTORE DEL DNA	3	30/09/2019	30/04/2020			
ISTR 01 PO 29 LIT TIPIZZAZIONE PCR - SSP	3	30/09/2019	01/10/2020			
ISTR 01 PO 30 LIT TIPIZZAZIONE PCR - SSO LUMINEX	3	30/09/2019	01/10/2020			
ISTR 01 PO 31 LIT MONITORAGGIO ATTECCHEMENTO KIT STR	3	30/09/2019	01/10/2020			
ISTR 01 PO 32 LIT TIPIZZAZIONE MEDIANTE PCR - SBT	3	30/09/2019	01/10/2020			
ISTR 01 PO 33 LIT TIPIZZAZIONE HLA MEDIANTE NGS	3	30/09/2019	29/04/2020	01/10/2020	25/09/2020	
ISTR 02 PO 24 LIT ESECUZIONE DEL VPPV TEST OPEN TUBE TEST	3	30/09/2019	01/10/2020			
ISTR 02 PO 25 LIT PREPARAZIONE DEI REAGENTI	3	30/09/2019				
ISTR 02 PO 26 LIT ESECUZIONE BACK - UP DATI	3	30/09/2019	09/04/2020			
ISTR 02 PO 27 LIT TIPIZZAZIONE HLA PER LA RICERCA DI UN DONATORE NON CORRELATO	3	30/09/2019	01/10/2020			
ISTR 02 PO 28 LIT QUANTIFICAZIONE DEL DNA	3	01/04/2020				
ISTR 02 PO 29 LIT GEL ELETROFORESI	3	30/09/2019	01/10/2020			
ISTR 02 PO 30 LIT SEPARAZIONE SOTTOPOPOLAZIONI CELLULARI	3	30/09/2019	01/10/2020			
ISTR 03 PO 31 LIT TIPIZZAZIONE HLA MEDIANTE NGS	3	30/09/2019	01/10/2020			
ISTR 03 PO 32 LIT DETERMINAZIONE E IDENTIFICAZIONE DI ANTICORPI ANTI - HLA	3	30/09/2019	non in uso			
ISTR 03 PO 33 LIT DETERMINAZIONE DI ANTICORPI anti-HLA MEDIANTE TEST DI SCREENING FlowPRA	3	01/04/2020	01/02/2021			
ISTR 03 PO 34 LIT DETERMINAZIONE DI ANTICORPI anti-HLA CON METODICA SCREENING LABscreen MIXED 1 E II	3	01/04/2020				
ISTR 04 PO 34 LIT IDENTIFICAZIONE DI ANTICORPI anti-HLA MEDIANTE TEST LUMINEX SINGLE ANTIGEN (LSA)	3	01/04/2020	01/02/2021			

Procedure e Istruzioni

Bambino Gesù OSPEDALE PEDIATRICO	PROCEDURA OPERATIVA Estrazione del DNA	Cod. PO 28 LIT
		Rev. 1
		Pagina 1 di 6

Data di emissione: 05/04/2023 Data di entrata in vigore: 19/04/2023

Bambino Gesù OSPEDALE PEDIATRICO	ISTRUZIONE ESTRAZIONE DEL DNA	Cod. ISTR.01 PO28 LIT
		Rev.0
		Pagina 1 di 5

Data di emissione: 22/09/2022 Data di entrata in vigore: 06/10/2022

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Approvato ed emesso: Dott. Marco Andreani
Responsabile LIT

Firma _____

Data: 22/09/2022

SOMMARIO

- 1 SCOPO E CAMPO DI APPLICAZIONE
- 2 RIFERIMENTI NORMATIVI
- 3 DEFINIZIONI E SIGLE
- 4 DESCRIZIONE DELLE ATTIVITA'
- 5 DOCUMENTAZIONE
- 6 RESPONSABILITA'
- 7 ALLEGATI

1. METODICHE DI ESTRAZIONE

1.1. METODICA DI ESTRAZIONE DI DNA DA SANGUE PERIFERICO, SANGUE MIDOLLARE E SOTTOPOPOLAZIONI LINFOCITARIE.

I prelievi di sangue periferico e sangue midollare e pellet delle sottopolazioni linfocitarie (precedentemente separate e diluite in 400 ml di PBS) vengono trattati direttamente con la metodica EZ1 ® ADVANCED XL QIAGEN, in base al Manuale utente EZ1 Advanced XL 05/2009 A-5.

1.4. METODICA DI ESTRAZIONE CON ESTRATTORE AUTOMATICO EZ1 ® ADVANCED XL QIAGEN

Reagenti e Materiale

- Estrattore EZ1 ® ADVANCED XL QIAGEN.
- Cartucce reagenti (da n. 1 a n. 14 a seconda del numero di campioni da estrarre) realizzate in polipropilene allungiate su un apposito supporto

Preparato: Sig.ra Annalisa Guagnano	Approvato ed emesso: Responsabile LIT _____ (Dott. Marco Andreani)
Verificato: Dott.ssa Nadia Tuccini Responsabile Funzione Certificazione	Data: 05/04/2023

Alcuni esempi di gestione della qualità

- Caratteristiche delle aree di lavoro
- Processo di accettazione dei campioni biologici
- Appropriatezza della documentazione
- **Appropriata manutenzione delle attrezzature**
- Controlli di qualità esterni
- Qualifica del Personale
- Validazione

Appropriata manutenzione delle attrezzature

Stabilire politiche e procedure per una corretta manutenzione delle apparecchiature

 Bambino Gesù OSPEDALE PEDIATRICO	PROCEDURA di PROCESSO TRASVERSALE Infrastrutture	Cod. Proc. PPT 7.1.3 LAB OPBG Rev. 1 Pagina 2 di 13
Data di emissione: 15/07/2023	Data di entrata in vigore: 30/07/2023	

Requisito ISO 7.1.3 – Infrastrutture

1. SCOPO E CAMPO DI APPLICAZIONE

La presente procedura definisce i compiti, le responsabilità e le modalità operative relative alla gestione e manutenzione delle infrastrutture dei Laboratori dell'Ospedale Pediatrico Bambino Gesù (LAB OPBG), quali locali, strumentazioni, hardware, software, sicurezza, etc., alla formazione del personale necessaria per l'acquisizione e il mantenimento delle competenze per conseguire la conformità e la qualità del servizio.

C2.2.1	The laboratory must establish and employ policies and procedures for the proper maintenance of equipment, instruments and test systems by:
C2.2.1.1	Defining its preventive maintenance programme for each instrument and piece of equipment at least once a year
C2.2.1.2	Performing and documenting function checks on equipment with at least the frequency specified by the manufacturer
C2.2.1.3	The laboratory must use calibrated dispensing instruments (e.g. pipettes, etc.) to perform assays
C2.2.1.3.1	Calibration of dispensing instruments must be performed at least once a year
C2.2.1.3.2	Calibration must be documented
C2.2.2	Refrigerators and freezers:
C2.2.2.1	Acceptable ranges for each refrigerator and freezer must be documented
C2.2.2.2	Must be monitored to detect unacceptable temperatures
C2.2.2.3	Should be coupled to recording thermometers
C2.2.2.4	Should be coupled to alarm systems with an audible alarm where it can be heard 24 hours a day
C2.2.2.5	Corrective actions for when the temperature is outside the documented acceptable range must be defined and documented

Appropriata manutenzione delle attrezzature

Calendario annuale che identifichi le date della manutenzione per ciascuna grande o piccola attrezzatura

C2.2.1	The laboratory must establish and employ policies and procedures for the proper maintenance of equipment, instruments and test systems by:
C2.2.1.1	Defining its preventive maintenance programme for each instrument and piece of equipment at least once a year
C2.2.1.2	Performing and documenting function checks on equipment with at least the frequency specified by the manufacturer

Inv. IC	Inv. OPBG	Matricola	Classe apparecchio	Produttore	Modello	Data Collaudo	Modalità acquisizione	Ultima Manut. Preventiva	prossima manutenzione
16396	110469	28715-1801	CAPPA ASPIRANTE	ERLAB DFS SA	Captair 481	27/08/2018	Proprietà	08/10/2024	08/10/2025
16283	110420	5382HG523229	AGITATORE DA LABORATORIO	EPPENDORF AG	THERMOMIXER C	06/09/2018	Proprietà	23/11/2023	22/11/2024
16085	110378	535533201	AGITATORE DA LABORATORIO	EPPENDORF AG	5355 THERMOMIXER COMFORT	28/07/2018	Proprietà	23/11/2023	22/11/2024
17396	128943	1904006494	LETTORE PER IMMUNOCHIMICA	TECAN AG	INFINITE M NANO PLUS	14/06/2019	Proprietà	20/08/2024	20/08/2025
16034	110304	272S3231377	AMPLIFICATORE DI SEQUENZE NUCLEOTIDICHE	APPLIED BIOSYSTEMS INC (LIFE TECHNOLOGIES)	GENE AMP PCR SYSTEM 2720	20/07/2018	Proprietà	19/10/2023	18/10/2024
21746		FM3DD21053021	ANALIZZATORE AUTOMATICO PER IMMUNOCHIMICA	LUMINEX CORP	LABSCAN 3D	01/06/2022	Comodato	12/07/2024	12/07/2025

Appropriata manutenzione delle attrezzature

**Report di manutenzione per
ciascuno strumento, inclusa
ovviamente ogni singola
pipetta automatica**

Calibration Report

Pipette Serial Number	A1551142T	Owner Company	SERVIZIO ASSISTENZA TECNICA
Pipette Second ID		Owner Department	Tel. 02.55404.333 - sat@eppendorf.it
Pipette Type	Pipet-Lite LTS L-1000 1ch	Owner Name	OSPED.BAMBIN GESU' - RM -
Manufacturer	Rainin	Method Description	
Method	Rainin Pipet-Lite L1000	3 Vol. 4 Meas. 100, 500, 1000µl	

Test Conditions		Balance	
Water Temperature	21,8 °C	Serial Number	1129140601
Humidity	60,2 %	Name	SAG285
Abs. Air Pressure	1008,3 hPa	Model	SAG285
Z-Factor	1.0033 µl/mg	Readability	0,0001 g
Z-Factor Reference	ISO 8655	Location	
Evaporation	0 µl/cycle	Tips	Puntali Rainin LTS 1000µl

As returned						
Weighings [g/µl]	100µl		500µl		1000µl	
	0,1006	100,93	0,4954	498,04	0,9929	996,18
	0,1005	100,83	0,4956	498,24	0,9884	991,66
	0,1003	100,63	0,4954	497,03	0,9945	997,78
	0,1009	100,63	0,4958	497,44	0,9944	997,68

Results [µl]	100µl		500µl		1000µl	
	Mean	Limits	Mean	Limits	Mean	Limits
Systematic Error [µl]	0,76	± 8,0	-2,31	± 8,0	-4,17	± 8,0
Systematic Error [%]	0,76		-0,46		-0,42	
Random Error [µl]	0,15	3,0	0,55	3,0	2,87	3,0
Random Error [%]	0,15		0,11		0,29	
Uncertainty meas. [µl]	1,06		3,42		9,92	
Status	Passed		Passed		Passed	
Status	Passed					

Date 16/11/2021

Performed by Mila Anghileri 

Notice

Cert.Tar.n. 7708184 Tested on SAG 285 s/n 1129140601 scad. 04/05/2022 C/N 0.005 a 200 g. INCERT.U 0.0000305 Rif ASOP P39 020-11/052019(salvo diversa specificata nel metodo)

100 µl, Systematic Error [µl]

500 µl, Systematic Error [µl]

Alcuni esempi di gestione della qualità

- Caratteristiche delle aree di lavoro
- Processo di accettazione dei campioni biologici
- Appropriatezza della documentazione
- Appropriata manutenzione delle attrezzature
- **Controlli di qualità esterni**
- Qualifica del Personale
- Validazione

Controlli di qualità esterni

Specifiche SOP che determinino le modalità di esecuzione dei Controlli di Qualità Esterni

SECTION D – EXTERNAL PROFICIENCY TESTING		
D1	PROCEDURE OF EXTERNAL PROFICIENCY TESTING	
D1.1	Registration for EPT schemes	
D1.1.1	<i>The laboratory must participate in EPT programme(s) to cover</i>	<input checked="" type="checkbox"/> <input type="checkbox"/>
D1.1.1.1	All the accredited laboratory applications (HLA typing, antibody screening and identification, crossmatching, etc.)	<input checked="" type="checkbox"/> <input type="checkbox"/>
D1.1.1.2	All techniques used individually or in combination as routinely employed to produce a final result	<input checked="" type="checkbox"/> <input type="checkbox"/>
D1.1.2	If no established scheme exists for a specific category (e.g. HNA antibody detection and identification) laboratory must participate in an EPT workshop or trial offered by an EPT Provider or must take part in an inter-laboratory exchange of samples	<input type="checkbox"/> <input checked="" type="checkbox"/> <input type="checkbox"/>
D1.1.3	If (an) EPT scheme(s) or EPT workshop(s)/trial(s) for a specific category exist(s) but the laboratory has no access, the laboratory must at least participate in an inter-laboratory exchange of samples.	<input checked="" type="checkbox"/> <input type="checkbox"/> <input type="checkbox"/>
D1.2	<i>The laboratory must prospectively define core and supplemental techniques according to the Accreditation Application.</i>	<input checked="" type="checkbox"/> <input type="checkbox"/>
D1.2.1	Core techniques are used individually or in combination to produce a final result	<input type="checkbox"/> <input checked="" type="checkbox"/> <input type="checkbox"/>
D1.2.2	Supplemental techniques are used occasionally for rare cases in combination with core techniques to refine final results	<input type="checkbox"/> <input checked="" type="checkbox"/> <input type="checkbox"/>
D1.3	<i>The laboratory must</i>	
D1.3.1	Prospectively document the relevant EPT schemes or workshops on an annual basis	<input checked="" type="checkbox"/> <input type="checkbox"/>
D1.3.2	Have a predetermined policy for testing EPT samples and must document this prior to the annual commencement of the EPT cycle	<input checked="" type="checkbox"/> <input type="checkbox"/>
D1.3.3	Have a predetermined policy if they select individual shipments or samples for EPT	<input type="checkbox"/> <input checked="" type="checkbox"/> <input type="checkbox"/>
D1.3.4	Have a predetermined policy for the selection of samples or shipments for supplemental techniques	<input type="checkbox"/> <input checked="" type="checkbox"/> <input type="checkbox"/>

Controlli di qualità esterni

 Bambino Gesù OSPEDALE PEDIATRICO	ISTRUZIONE CQ ESTERNI/ CQ INTERNI		Cod. ISTR.04 GEN – LIT
		Rev.1	
Data di emissione:	14/02/2023	Data di entrata in vigore:	01/03/2023

Approvato ed emesso: Dott. Marco Andreani

Responsabile LIT

Firma _____ Data: _____

1. CONTROLLI DI QUALITA' ESTERNI

Il LIT si impegna a partecipare ai Controlli di qualità Esterni in base agli Standard EFI.

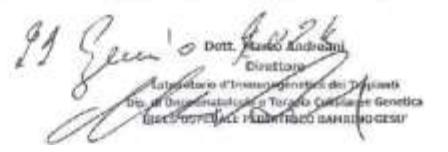
- **Tipizzazione molecolare HLA:** Il LIT partecipa ad un programma di qualità organizzato dall'Istituto Superiore di Sanità (ISS) per quanto riguarda la tipizzazione HLA a bassa ed alta risoluzione.
- **Tipizzazione molecolare KIR:** Il LIT partecipa ad un programma di interscambio laboratori per quanto riguarda la tipizzazione dei geni KIR.

- **Ricerca di anticorpi anti-HLA:** Il LIT partecipa sia per lo screening che per la ricerca delle singole specificità anticorpali ai seguenti EPT:
 - UK NEQAS programma di qualità
 - ASHI programma di qualità
 - Intercambio laboratori per quanto riguarda lo Screening and identificazione IgM organizzato dall'ASHI
- **Crossmatch citofluorimetrico (FC-XM):** Il LIT partecipa al programma di qualità organizzato UK NEQAS.
- **Crossmatch con metodica di citotossicità Complemento-Dipendente (CDC-XM):** Il LIT partecipa al programma di qualità organizzato UK NEQAS.
- **Test anti-HLA citotossici Clq, Test recettore AT1R dell'Angiotensina:** Il LIT partecipa al programma interscambio di qualità organizzato ASHI
- **Monitoraggio post-trapianto di midollo allogenico:** Il LIT partecipa al programma di EPT gestito da ISS.

Controlli di qualità esterni

Calendario specifico per l'esecuzione dei Controlli di Qualità Esterni per la tipizzazione HLA molecolare, suddiviso per numero di campioni biologici da testare e le metodiche da utilizzare

Provider CQ ISS-2024	LOW RESOLUTION					HIGH RESOLUTION								Cod.: MD 03 ISTR 04 GEN-LIT	
	HLA-A	HLA-B	HLA-C	HLA-DRB1	HLA-DQB1	HLA-A	HLA-B	HLA-C	HLA-DRB1	DRB3-DRB4-DRB5	HLA-DQA1	HLA-DQB1	HLA-DRB1	HLA-DPA1	
1° INVIO 13-02-2024															
Campione 1	SSO-XB-NGS SSP	SSO-XB- NGS	SSO-XB- NGS	SSO-XB-NGS	SSO-XB-NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	
Campione 2	SSO-XB-NGS	SSO-XB- NGS SSP	SSO-XB- NGS	SSO-XB-NGS	SSO-XB-NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	
Campione 3	SSO-XB-NGS	SSO-XB- NGS	SSO-XB- NGS SSP	SSO-XB-NGS	SSO-XB-NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	
Campione 4	SSO-XB-NGS	SSO-XB- NGS	SSO-XB- NGS SSP	SSO-XB-NGS	SSO-XB-NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	
2° INVIO 07-05-2024															
Campione 5	SSO-XB-NGS	SSO-XB- NGS	SSO-XB- NGS	SSO-XB-NGS	SSO-XB- SSP	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	
Campione 6	SSO-XB-NGS	SSO-XB- NGS	SSO-XB- NGS	SSO-XB-NGS	SSO-XB- SSP	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	
Campione 7	SSO-XB-NGS	SSO-XB- NGS SSP	SSO-XB- NGS	SSO-XB-NGS	SSO-XB- SSP	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	
3° INVIO 17-09-2024															
Campione 8	SSO-XB-NGS	SSO-XB- NGS SSP	SSO-XB- NGS	SSO-XB-NGS	SSO-XB- SSP	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	
Campione 9	SSO-XB-NGS	SSO-XB- NGS	SSO-XB- NGS SSP	SSO-XB-NGS	SSO-XB- SSP	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	
Campione 10	SSO-XB-NGS	SSO-XB- NGS	SSO-XB- NGS SSP	SSO-XB-NGS SSP	SSO-XB- NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	SSO-XB/ NGS	


Dott. Paolo Andreatta
Dirigente
Istituto d'Immunogenetica del Trapianto
Istituto Nazionale di Tumori - Centro Genetica
Irc-Cnr - Iasi - Istituto Paediatrico Bambino Gesù

Controlli di qualità esterni

D1.5	Minimum number of samples for EPT per year			
D1.5.1	<i>The minimum number of samples applies to all core techniques used to produce a final result:</i>			
D1.5.1.1	Serological typing: 10 samples	✓		
D1.5.1.2	Each low resolution DNA-based typing technique: 10 samples		✓	
D1.5.1.3	Each high resolution DNA-based typing technique: 10 samples		✓	
D1.5.1.4	Each allelic resolution DNA-based typing technique: 10 samples	✓		
D1.5.1.5	HPA/HNA/KIR/MICA typing: 10 samples		✓	
D1.5.1.6	HLA antibody detection: 10 samples for HLA class I and 10 samples for HLA class II The same samples can be used for the detection of both classes		✓	
D1.5.1.7	HLA antibody identification by CDC: 10 samples		✓	
D1.5.1.8	HLA antibody identification by solid phase assays: All HLA class I and II antibody positive samples as defined in D1.5.1.6. If the HLA antibody identification is a separate scheme the minimum number is 10 samples. A laboratory may test only for class I or class II antibodies according to their clinical requirements and D1.1		✓	
D1.5.1.9	HPA/MICA antibody detection and identification: 5 samples	✓		
D1.5.1.10	Crossmatching: 20 tests of different donor/recipient combinations of each accredited cell subtype (B-/T-/unseparated cells) which must include a minimum of two cell samples and 10 different sera		✓	
D1.5.1.11	Haematopoietic chimaerism and engraftment monitoring: 10 tests of different donor/recipient mixtures in the range 0% - 100% excluding the reference donor and recipient samples		✓	

Specifici standard di riferimento relativi al numero minimo di campioni biologici da testare per ciascuna singola metodica nei Controlli di Qualità Esterni

Alcuni esempi di gestione della qualità

- Caratteristiche delle aree di lavoro
- Processo di accettazione dei campioni biologici
- Appropriatezza della documentazione
- Appropriata manutenzione delle attrezzature
- Controlli di qualità esterni
- **Qualifica del Personale**
- Validazione

Qualifica del Personale

SECTION B – PERSONNEL QUALIFICATIONS

B1	For the purposes of this document, EFI defines the Director as the person who is responsible for the H&I laboratory
B2	The laboratory must employ one or more individuals who meet the qualifications and fulfil the responsibilities of:
B3	The Director and/or Co-Director
B3.1	A Director, that must:
B3.1.1	Hold a qualification approved by EFI, such as an ESHI or national diploma, earned doctoral degree in a biological science, or be a physician, and
B3.1.2	Have minimum qualifying experience equivalent to either of the following:
B3.1.2.1	Four years' relevant experience two of which were devoted to full time training in human H&I testing, or
B3.1.2.2	Four years of working experience at full time in human H&I testing
B3.1.2.3	Additional qualifications required according to national legislation also apply
B3.1.3	Have documentation of professional competence in the appropriate activities in which the laboratory is engaged. This should be based on sound knowledge of the fundamentals of immunology, genetics and histocompatibility testing
B3.1.4	If a Co-Director is appointed, this person must also fulfil Standards B1.1.1 - B1.1.3
B3.1.5	The Director and/or Co-Director must:
B3.1.5.1	Be available on site to supervise the laboratory for at least 80% of the week
B3.1.5.2	Provide adequate supervision of technical personnel
B3.1.5.3	Utilises his/her special scientific skills in developing new procedures
B3.1.5.4	Be held responsible for the proper performance, interpretation and reporting of all laboratory procedures

Qualifica del Personale



**European Federation for Immunogenetics
Accreditation Programme**

APPLICATION FOR EFI DIRECTORSHIP

Every applicant for directorship according to EFI standards B must provide the information requested on this form and the additional documents as listed at the end of this form. Please send the completed form and the documents to your Regional Commissioner.

Clinical categories of the applicant's laboratory (please check boxes):

- Renal and/or pancreatic transplantation
- Other solid organ transplantation (organs: _____)
- Haematopoietic stem cell transplantation
 - Donor registry typing
 - Related stem cell transplantation
 - Unrelated stem cell transplantation
 - Cord blood transplantation
- Transfusion
- Disease association
- Chimaerism



**European Federation for Immunogenetics
Accreditation Programme**

Table 1: Experience and expertise in molecular immunogenetics

Only samples/examinations for which a technical and medical plausibility check and validation have been carried out are counted. Please specify approximate number of tests for each method. If number exceeds 1000, specifying ">1000" is sufficient.

Technical Category	Approx. number of tests	Methods
Low-resolution HLA typing (minimum HLA-A, -B, -C, -DRB1 and -DQB1 per sample)		
High-resolution HLA typing (minimum HLA-A, -B, -C, -DRB1 and -DQB1 per sample)		
Selective HLA typing (e.g. HLA-B27)		HLA antigen or allele/resolution of typing/clinical application:

Qualifica del Personale



European Federation for Immunogenetics Accreditation Programme

Table 3: Experience and expertise in risk assessment and consulting. If number exceeds 1000, specifying ">1000" is sufficient.

Risk assessment and consulting for renal and/or pancreatic transplantation	Approx. number of living donor transplants:	Approx. number of deceased donor transplants:
Risk assessment and consulting for other organ transplantation	Approx. number of transplants:	Organs:
Risk assessment and consulting for stem cell transplants	Approx. number of related donor transplants:	Approx. number of unrelated donor transplants:
Risk assessment and consulting for transfusion	Approx. number:	Transfusion product:
Risk assessment and consulting for disease association	Approx. number:	Diseases:
Risk assessment and consulting for chimaerism	Approx. number	

The following documents must be provided (please check boxes):

- Curriculum vitae** (clearly define education e.g. biologist and PhD in biological science)
- Description of duties in the lab**
- List of publications**
- List of continuing education** (for format please see EFI Accreditation Application Packet: organiser, meeting/title of lecture, duration, level of participation)
- ESHI diploma and/or national/international H&I specialisation diploma (if applicable)**

- I confirm that all information is truthful and accurate.
- I consent to the storage and use of the data submitted in this application for the purposes of EFI accreditation.
- Upon request by the accreditation committee, I agree to an oral discussion about my stated experience and expertise.

Date, signature of the applicant

Alcuni esempi di gestione della qualità

- Caratteristiche delle aree di lavoro
- Processo di accettazione dei campioni biologici
- Appropriatezza della documentazione
- Appropriata manutenzione delle attrezzature
- Controlli di qualità esterni
- Qualifica del Personale
- **Validazione**

Validazione nuove metodiche

Il Laboratorio deve disporre di strumenti utili al monitoraggio delle attività

Il Laboratorio deve validare/verificare qualsiasi nuovo test prima che questo venga introdotto nella routine

C1.3	Systems for Continuous Test Evaluation and Monitoring
C1.3.1	The laboratory must establish and employ policies and procedures, and document actions taken when:
C1.3.1.1	Test systems do not meet the laboratory's established criteria
C1.3.1.2	Quality control results are outside of acceptable limits
C1.3.1.3	Errors are detected in the reported patient results. In this instance, the laboratory must:
C1.3.1.3.1	Promptly notify the authorised person ordering or individual utilising the test results of reporting errors
C1.3.1.3.2	Issue corrected reports
C1.3.1.3.3	Maintain copies of the original report as well as the corrected report for at least two years
C1.3.2	The laboratory must have mechanisms in place for continuous monitoring of all test systems and equipment used, including:
C1.3.2.1	Validation/verification, before introduction into routine use, of all new tests, by systematic comparative evaluation of results obtained in parallel with the new and the standard system
C1.3.2.2	Regular evaluation of results obtained in external and internal QC testing
C1.3.2.3	Regular monitoring of test validity in routine testing, by recording observations diverging from the expected results (e.g. cross-reactivity of probes or primer mixes, day-to-day variations)

Validazione nuove metodiche

Evidenza scritta di confronto tra i risultati ottenuti con metodiche precedentemente utilizzate in laboratorio o con panel di riferimento

 Bambino Gesù OSPEDALE PEDIATRICO	VALIDAZIONE METODICHE, SOFTWARE E STRUMENTAZIONE	Cod.: MD 05 PO22 LIT
Data di emissione:	23/03/2023	Rev. 0
	Data di entrata in vigore:	Pagina 1 di 1

Dal 25/09/2023 al 03/10/2023 il Laboratorio di Immunogenetica dei trapianti ha eseguito la validazione dell'estrattore automatico EZ2 – Connect MDx.

La validazione della metodica ha confermato le seguenti indicazioni:

- Formazione per i quattro tecnici addetti all'accettazione (4 ore)
- Estrazione di 20 campioni di sangue periferico e midollare in doppio su piattaforma EZ-1 Advanced XL e EZ-2 Connect MDx
- Confronto della purezza e della quantità del DNA ottenuto da entrambe le piattaforme
- Tipizzazione HLA per 11 loci con metodica NGS di 6 dei 20 campioni di DNA estratti sia da EZ-1 Advanced XL che da EZ-2 Connect MDx per verificare la congruità dei risultati
- Caratterizzazione di STR di 2 dei 20 campioni di DNA estratti sia da EZ-1 Advanced XL che da EZ-2 Connect MDx per 16 sistemi con il kit identifier

SI NO

Pertanto a partire dal 04/10/2023 l' EZ2 – Connect MDx può essere utilizzato nella routine del laboratorio.

Roma, 04/10/2023

Operatore



Responsabile


Dott. Marco Andreani
Direttore

Laboratorio d'Immunogenetica dei Trapianti
Dip. di Oncematologia e Terapia Cellulare e Genetica
IRCCS OSPEDALE PEDIATRICO BAMBINO GESÙ

Validazione nuove metodiche

Quanti test per validare una nuova metodica....?

Work flow - NGS	
Platform	XY, YX etc.
Kit used	Name
Number of samples to test	20 – 60 up to 100
Number of alleles to test	100 - 300 up to 500

Validazione nuovi reagenti

**Evidenza scritta di validazione per
ogni nuovo reagente venga
introdotto in laboratorio prima del
suo utilizzo nella routine**

	VERBALE DI VALIDAZIONE KIT HLA	Cod.: MD 02 PO 25 LIT
		Data: 30/09/2019
		Ed. 3 – Rev. 0
		Pagina 1 di 1

IDENTIFICAZIONE DEL KIT

Descrizione	Ditta	Temp.	Lotto	Scadenza	Spedizione	Data arrivo

IDENTIFICATIVO VALIDAZIONE

Data	DNA Rif.N°	Esito Validazione	Firma

INIZIO UTILIZZO:

OSSERVAZIONI	ANALISI DEL DATO

FIRMA RESP.:data.....

Validazione EPT esterni

Il Laboratorio deve disporre di strumenti utili al monitoraggio delle attività

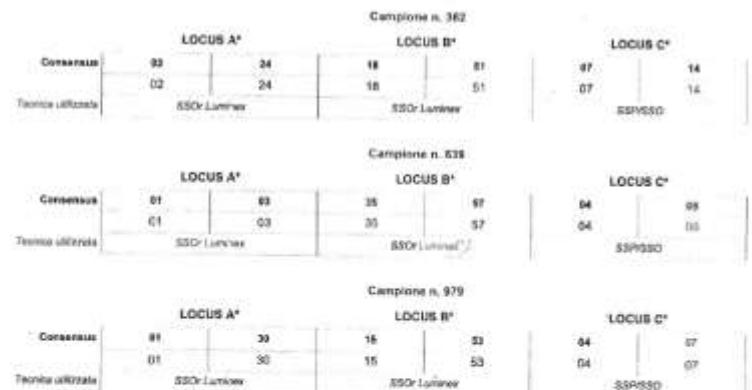
Il Laboratorio deve valutare con regolarità i risultati relativi ai controlli di qualità esterni ed interni

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Validazione EPT esterni

Italian National EPT in Immunogenetics
Tipizzazione genomica HLA (Bassa Risoluzione)
Report Consensus - Invio n° 3 - Anno 2024

Laboratorio: ROMA OPBG - CLASSE I DNA (Bassa Risoluzione)



Risultati condivisi sullo
zincico settivo L.1 dal 9/10/24

Dott. Marco Andreani
Coordinatore
Laboratorio di Immunogenetica del Trapianto
C/o di Oncogenetologia e Terapia Cellulare e Genetica
IRCCS OSPEDALE PEDIATRICO BAMBINO Gesù

Bambino Gesù OSPEDALE PEDIATRICO		RIUNIONE DI FORMAZIONE INFORMAZIONE		Cod.: MD 02 ISTR 01 GEN-LIT	
				Rev. 0	
				Pagina 1 di 1	
				Data di emissione: 23/03/2023 Data di entrata in vigore: 06/04/2023	
				Titolo riunione: Riunione 1/2 Data: 4/10/24	
				Ora inizio riunione: 11:30 Ora fine riunione: 13:30	
Partecipanti		Partecipanti		firma	
Andreani Marco Battarra Manarosa Troiano Maria Galluccio Tiziana Giustiniani Paola Blanculli Antonio Giuseppe Testa Giuseppe		Guagnano Annalisa Di Luzio Andrea Besi Francesca Mangiono Martina		firma	
Valutazione delle non conformità (NC)					
Numerazione e tipo di NC		N. _____ Cat. _____ Descrizione: _____		N. _____ Cat. _____ Descrizione: _____	
		N. _____ Cat. _____ Descrizione: _____		N. _____ Cat. _____ Descrizione: _____	
Azione Correttiva NC		N. _____		N. _____	
		N. _____		N. _____	
Verifica risoluzione NC		N. _____		N. _____	
Valutazione CQ esterni		data: 9/10/24 ISS HLA L.R.: <u>concordanza</u> C.R. <u>corretto</u> data: 9/10/24 ISS HLA H.R.: <u>risultato corretto</u> C.R. <u>corretto</u> data: 9/10/24 ISS Attecchimento: <u>risultato corretto</u> C.R. <u>corretto</u> data: 9/10/24 Interscambio Laboratori KIR: <u>risultato corretto</u> C.R. <u>corretto</u> data: 9/10/24 UK NEQASS anticorpi anti-HLA: <u>risultato corretto</u> C.R. <u>corretto</u> data: 9/10/24 ASHI anticorpi anti-HLA: <u>risultato corretto</u> C.R. <u>corretto</u>			
Valutazione CQ interni		CQ interno eseguito da: _____ data: 9/10/24 risultato: _____ N.F. CORRETTO			
Aggiornamento accreditamento. EPI					
Wipe Test e/o open tube test		Eseguito da: <u>test</u> data: 9/10/24 risultato: <u>N.F.A.V.V.</u> <u>corretto</u>			

Validazione EPT esterni

Il Laboratorio deve disporre di strumenti utili al monitoraggio delle attività

Il Laboratorio deve valutare con regolarità i risultati relativi ai controlli di qualità esterni ed interni

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Validazione risultati non attesi

Il Laboratorio deve disporre di strumenti utili al monitoraggio delle attività

Il Laboratorio deve avere evidenza scritta di risultati non attesi ottenuti durante la normale attività di routine

		Monitoraggio Qualità NGS							Cod.: MD 02 PO 33 LIT Rev. 0	Pag. 1
		Data di Emissione: 23/03/2023			Data di entrata in vigore: 06/04/2023					
Amp	KIT	Lotto	scadenza	Ditta	data	#DNA	Cod Anomali	Natura dell'anomali	Note	
1	FASTPLEX	4	13/2023	onelambda	13/03/2023	0742-2023		LOW COVERAGE DRB3/4/5	NO DRB3/4/5	
1	FASTPLEX	4	28/03/2023	onelambda	21/03/2023	0903-2023		POLIG		
1	FASTPLEX	4	28/03/2023	onelambda	28/03/2023	1003-2023				
6	FASTPLEX	4	giu-24	onelambda	19/10/2023	3217-2023	7	A*11:21N - FARE SSO		
6	FASTPLEX	4	giu-24	onelambda	19/10/2023	3218-2023	7	A*01:04:01N FARE SSO		
14	FASTPLEX	4	giu-24	onelambda	14/11/2023	3447-2023	7	ERRORE DQA1/DRB3		

Documenti utili



Associazione Italiana di Immunogenetica e Biologia dei trapianti

Linee-Guida AIBT per la Valutazione dell'Istocompatibilità nel Trapianto d'Organo

In sinergia con la Società Italiana Trapianti d'Organo (SITO)

(Versione 06/07/2016)

Received: 26 April 2018 | Accepted: 28 April 2018
DOI: 10.1111/wim.13289WILEY HLA
Human Leukocyte Antigen

REVIEW ARTICLE

Accreditation of histocompatibility and immunogenetics laboratories: Achievements and future prospects from the European Federation for Immunogenetics Accreditation Programme

A. Harmer^{1,2} | L. Mascaretti^{2,3} | E. Petershofen^{2,4}

Associazione Italiana di Immunogenetica e Biologia dei trapianti

RACCOMANDAZIONI AIBT PER LA VALUTAZIONE DELLA ISTOCOMPATIBILITÀ NEL TRAPIANTO DI CELLULE STAMINALI EMOPOIETICHE

(Versione 1.1 del 20 Luglio 2022)

Andreani,Marco
Crocchiolo, Roberto
Falco,Michela
Fusco,Caterina
Garbarino,Lucia
Papola,Franco
Rombolà,Gianni
Vecchiatto,Cinzia

Approvato dal Consiglio Direttivo AIBT in data 21/01/2022

Presidente Dr. Franco Papola
Vice Presidente Dr. Giovanni Rombolà
Segretario Dr. Roberto Crocchiolo
Tesoriere Dr.ssa Lia Mele
Consigliere Dr.ssa Benedetta Allegra Mazzi

Condiviso con il Gruppo Italiano per il Trapianto di Midollo Osseo, cellule staminali emopoietiche e terapia cellulare (GITMO) in data 06/04/2022.

Presidente: Prof. Fabio Ciceri

Gruppo degli Ispettori EFI



Per informazioni: la
segreteria EFI



<http://www.efiweb.eu/>

[http://www.efiweb.eu/efi-
committees.html](http://www.efiweb.eu/efi-committees.html)

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Grazie per l'attenzione

Laboratorio di Immunogenetica dei Trapianti (LIT)



Maria Troiano



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Antonio Bianculli



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Paola Giustiniani



Martina Mangione



Andrea Di Luzio



Giuseppe Testa



Annalisa Guagnano



Francesca Besi



Prof. Franco Locatelli